

# Linear Least-Squares Method for Unbiased Estimation of $T_1$ From SPGR Signals

Lin-Ching Chang,<sup>1,2\*</sup> Cheng Guan Koay,<sup>2</sup> Peter J. Basser,<sup>2</sup> and Carlo Pierpaoli<sup>2</sup>

**The longitudinal relaxation time,  $T_1$ , can be estimated from two or more spoiled gradient recalled echo images (SPGR) acquired with different flip angles and/or repetition times (TRs). The function relating signal intensity to flip angle and TR is nonlinear; however, a linear form proposed 30 years ago is currently widely used. Here we show that this linear method provides  $T_1$  estimates that have similar precision but lower accuracy than those obtained with a nonlinear method. We also show that  $T_1$  estimated by the linear method is biased due to improper accounting for noise in the fitting. This bias can be significant for clinical SPGR images; for example,  $T_1$  estimated in brain tissue (800 ms <  $T_1$  < 1600 ms) can be overestimated by 10% to 20%. We propose a weighting scheme that correctly accounts for the noise contribution in the fitting procedure. Monte Carlo simulations of SPGR experiments are used to evaluate the accuracy of the estimated  $T_1$  from the widely-used linear, the proposed weighted-uncertainty linear, and the nonlinear methods. We show that the linear method with weighted uncertainties reduces the bias of the linear method, providing  $T_1$  estimates comparable in precision and accuracy to those of the nonlinear method while reducing computation time significantly. Magn Reson Med 60:496–501, 2008. © 2008 Wiley-Liss, Inc.**

**Key words:**  $T_1$  estimation; linear model; accuracy; uncertainties; SPGR signals

Clinical imaging of the longitudinal relaxation time,  $T_1$ , in human subjects has several potential applications, including perfusion imaging (1), dynamic contrast imaging (2), assessment of Parkinson's disease (3), assessment of schizophrenia (4) and multiple sclerosis (5), and quantification of myocardial blood flow (6). In spite of its potential clinical utility, quantitative  $T_1$  mapping is not routinely used due to the long scanning time inherent in inversion recovery sequences. Recently, however, the acquisition of high-resolution  $T_1$  maps in a clinically feasible time frame has been demonstrated with Driven Equilibrium Single Pulse Observation of  $T_1$  (DESPOT1) (7,8). DESPOT1 derives  $T_1$  from two or more spoiled gradient recalled echo (SPGR) images acquired with a constant TR and different flip angles. The  $T_1$  maps have been computed on a voxel-by-voxel basis using linear least squares (LLS) fitting of a linear transformation of the function relating signal intensity, flip angle, TR,  $T_1$ , and equilibrium longitudinal mag-

netization,  $M_0$ . This transformed LLS fitting method—first described by Gupta (9) in 1977, which we denote as Gupta's LLS (GLLS), and used in many previous works (7,8,10–12)—has the advantage of being computationally efficient. However, our preliminary study found that the estimated  $T_1$  using GLLS was generally biased and overestimated (13).

In this work, we study systematically the bias in  $T_1$  computed from SPGR signals when different fitting methods are used. Monte Carlo simulations provide a detailed evaluation of the accuracy of  $T_1$  using GLLS and nonlinear least squares (NLS) methods in several experimental conditions. We also evaluate the performance of an intensity-based weighted LLS approach (ILLs) proposed by Deoni et al. (14). The ILLs method assigns greater weight to high signal-to-noise ratio (SNR) points in order to increase the precision of estimated  $T_1$ . Finally, we propose a new LLS approach that uses weighted uncertainties in the fitting (WLLS). The proposed WLLS method weights each image with the uncertainty that takes into account the adjustment of noise contribution due to the rearrangement of a nonlinear model into a linear one. Numerical and human brain data simulations are used to compare the accuracy of  $T_1$  estimates using the GLLS, ILLs, WLLS, and NLS methods.

## THEORY

### NLS Method

The measured SPGR signal intensity,  $s_i$ , is a function of the longitudinal relaxation time,  $T_1$ ; the repetition time (TR); the flip angle,  $\alpha_i$ ; and the equilibrium longitudinal magnetization,  $M_0$ :

$$s_i = \frac{M_0 \left( 1 - \exp\left(-\frac{TR}{T_1}\right) \right) \sin(\alpha_i)}{1 - \exp\left(-\frac{TR}{T_1}\right) \cos(\alpha_i)}. \quad [1]$$

The NLS approach estimates  $T_1$  and  $M_0$  from Eq. [1] by minimizing the following  $\chi^2$  objective function:

$$\chi_{\text{NLS}}^2(M_0, T_1) = \sum_{i=1}^n \frac{1}{\sigma_i^2} \left( s_i - M_0 \frac{1 - \exp\left(-\frac{TR}{T_1}\right) \sin(\alpha_i)}{1 - \exp\left(-\frac{TR}{T_1}\right) \cos(\alpha_i)} \right)^2, \quad [2]$$

where  $\sigma_i$  is the expected signal standard deviation (SD) due to noise. When the SNR,  $s_i/\sigma_i$ , is greater than 5,  $\sigma_i$  can be considered as a constant in all images, i.e.,  $\sigma_1 = \sigma_2$

<sup>1</sup>Department of Electrical Engineering and Computer Science, Catholic University of America, Washington, DC, USA.

<sup>2</sup>National Institute of Child Health and Human Development, National Institutes of Health, Bethesda, Maryland, USA.

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\*Correspondence to: Lin-Ching Chang, Catholic University of America, 620 Michigan Ave. NE, Washington, DC 20064. E-mail: changl@cua.edu

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= ... =  $\sigma_n = \sigma$ . But when  $s_i/\sigma_i$  is less than 5, this assumption becomes invalid (15).

### LLS Method

In the LLS method, the linear equation is obtained by assuming that TR is a constant. By rearranging Eq. [1] and denoting  $\exp(-TR/T_1)$  by  $E_1$ , we have:

$$\frac{s_i}{\sin(\alpha_i)} = E_1 \frac{s_i}{\tan(\alpha_i)} + M_0(1 - E_1). \quad [3]$$

Equation [3] can be rewritten in a more explicit linear notation as  $\mathbf{y} = \mathbf{b}\mathbf{x} + a$  with  $y_i = f(s_i) = s_i/\sin(\alpha_i)$ ,  $x_i = g(s_i) = s_i/\tan(\alpha_i)$ ,  $b = E_1$ , and  $a = M_0(1 - E_1)$ .

The slope,  $b$ , and the y-intercept,  $a$ , can be estimated by linear regression (16,17), i.e., by minimizing the  $\chi^2$  objective function:

$$\begin{aligned} \chi_{LLS}^2(a, b) &= \sum_{i=1}^n w(i)x(y_i - bx_i - a)^2 \\ &= \sum_{i=1}^n w(i)x \left( \frac{s_i}{\sin(\alpha_i)} - E_1 \frac{s_i}{\tan(\alpha_i)} - M_0(1 - E_1) \right)^2. \end{aligned} \quad [4]$$

The GLLS method proposed by Gupta (9) uses the linearization approach of Eq. [3] with  $w_{GLLS}(i) = 1/\sigma_i^2$ , which has been used in many previous works (7,8,10–12). Note that  $\sigma_i$  is the SD of the measurement error on  $y_i$  and is generally assumed to be a constant in a linear model when measurements of  $y_i$  are independent. The assumption in the widely-used GLLS method that the SD of  $y_i$  is the same for all data points (i.e.,  $\sigma_i = \sigma$ ) is incorrect because the experimental errors are distorted by the nonlinear to linear transformation. Under this assumption,  $T_1$  and  $M_0$  can then be calculated with the representation of  $a$  and  $b$  (10,16):  $T_1 = -TR/\ln b$  and  $M_0 = a/(1 - b)$ .

An empirical weighting approach for the LLS fitting was introduced in Ref. 14. This method, which we call the ILLS method, assigns greater weight to data points with higher signal intensity in an attempt to increase the precision of estimated  $T_1$ , and solves Eq. [3] by minimizing Eq. [4] with a different weighting function defined in Ref. 14:  $w_{ILLS}(i) = s_{\alpha_i}/s_{\alpha_E}$ , where  $s_{\alpha_i}$  is the signal intensity acquired with flip angle  $\alpha_i$ , and  $s_{\alpha_E}$  is the signal intensity acquired with the Ernst angle.  $T_1$  and  $M_0$  can be calculated as above.

Experimental errors are generally distorted when a nonlinear model is transformed to a linear one. A weighting function that accommodates the distorted uncertainties can be derived by using the error propagation theory (16) (see Appendix). Here, we can rewrite the  $\chi^2$  objective function for the NLS approach (Eq. [2]) as:

$$\chi_{NLS}^2 = \sum_{i=1}^n \frac{1}{\sigma_i^2} \left( s_i - M_0 \sin(\alpha_i) \frac{1 - E_1}{1 - E_1 \cos(\alpha_i)} \right)^2$$

$$= \sum_{i=1}^n \frac{1}{\sigma_i^2} \left( \frac{\sin(\alpha_i)}{1 - E_1 \cos(\alpha_i)} \right)^2 \left( \frac{s_i}{\sin(\alpha_i)} - E_1 \frac{s_i}{\tan(\alpha_i)} - M_0(1 - E_1) \right)^2. \quad [5]$$

Notice that Eq. [5] is similar in form to the objective function in LLS (i.e., Eq. [4]), and can be considered as a new weighted LLS method, which we denote as WLLS. By comparing Eq. [4] with Eq. [5], the weighting function of WLLS can therefore be defined as  $w_{WLLS}(i) = \frac{1}{\sigma_i^2} \left( \frac{\sin(\alpha_i)}{1 - E_1 \cos(\alpha_i)} \right)^2$ .

Here, we have established the theoretical connection between the nonlinear and proposed weighted linear methods based on the approach used in Ref. 18. The proposed WLLS method is equivalent to the NLS method in principle because Eq. [5] is derived directly from Eq. [2], and no transformation or approximation is applied during the rearrangement.

We now want to minimize Eq. [5] with respect to  $a$  and  $b$ . However, the occurrence of  $b (= E_1)$  in the denominator of Eq. [5] makes the task of fitting considerably harder. An iterative least-squares fitting approach can be used here to find the optimal solution (19), and the GLLS method can be used as an initial solution. There are strategies for fitting a line in this situation (17,20,21), any of which can be applied here. We use Brent's method as described in Refs. 22 and 23, which is a reasonable strategy for minimizing a general one-dimensional function so that the minimization with respect to  $b$  is also minimized with respect to  $a$ .

## MATERIALS AND METHODS

### Numerical Simulations

We evaluated the accuracy and precision of different fitting algorithms in several experimental conditions by performing Monte Carlo simulations. Noise-free SPGR signals were generated using Eq. [1] given a fixed TR value, multiple flip angles, a single expected value of the equilibrium longitudinal magnetization  $M_0$ , and various expected values of  $T_1$ . Different SNR ( $SNR_0$ ) levels were simulated by adding Gaussian noise ( $s_{noise}$ ) in quadrature with zero mean and variable SD,  $\sigma$ , to the noise-free SPGR signals, i.e.,  $s_{spgr} = \sqrt{(s_{noise-free-spgr} + s_{noise^1})^2 + s_{noise^2}^2}$  (24). The noise level is defined as  $M_0$  divided by the signal SD, i.e.,  $SNR_0 = M_0/\sigma$ . We calculated  $T_1$  by fitting the synthetic SPGR signals using the GLLS, WLLS, and NLS fitting approaches. The results reported in the next section are computed with TR = 10 ms,  $M_0 = 3000$ , and  $T_1 = 600, 800, 1000, 1200, 1600, \text{ or } 2000$  ms. The two optimal flip angles for the chosen  $T_1$  were computed according to the formula in Refs. 10 and 14. For example, given  $T_1 = 1000$  ms, TR = 10 ms, and  $M_0 = 3000$ , the optimal flip angles are 3.35° and 19.38°. Six SPGR images were used in simulations with repeated experiments of the two optimal angles without averaging. Noise levels were tested with  $SNR_0$  ranging from 30 to 300. Each set of parameters was repeated 131,072 times.

## Human Brain Simulation

A separate set of simulations was aimed at assessing the accuracy of estimated  $T_1$  for experimental conditions compatible with clinical studies of the human brain at 1.5T. We collected a high-quality SPGR dataset of the brain of a healthy male volunteer. Images were acquired with a DESPOT1 sequence (7) in axial view with  $0.9375\text{-mm}^2$  in-plane resolution, 2-mm slice thickness,  $TR = 8$  ms, and 20 different flip angles. The scan time was about 20 min (1 min per flip angle). This high-quality SPGR dataset was used to compute a  $T_1$  and an  $M_0$  map using the NLS method, and the values obtained in each voxel of the brain were assumed to be error-free. From these “gold standard”  $T_1$  and  $M_0$  maps, and a given set of TR and flip angles, we then generated synthetic noise-free SPGR images by computing the signal intensity in each voxel using Eq. [1]. Gaussian noise in quadrature with zero mean and a given SD,  $\sigma = 100$ , was added to the noise-free synthetic SPGR images as described in the previous paragraph. In the resultant SPGR brain images that have  $SNR_0 = M_0/\sigma$  ranging from 90 to 150 throughout most areas of the brain tissue, we then computed the  $T_1$  map by fitting these synthetic human brain data with GLLS, WLLS, and ILLS methods. This procedure was repeated 500 times and an averaged  $T_1$  map was created by taking the mean value of  $T_1$  from the 500 repeats on a voxel-by-voxel basis. The results reported in the next section are computed using  $TR = 10$  ms and six SPGR images, with the flip angles in each image equal to  $3^\circ$ ,  $6^\circ$ ,  $9^\circ$ ,  $12^\circ$ ,  $15^\circ$ , and  $18^\circ$ , respectively.

## RESULTS

Figure 1 demonstrates the behavior of  $T_1$  bias when estimating  $T_1$  from SPGR signals using the GLLS method. Figure 1a shows that the distributions of  $T_1$  obtained with GLLS are biased, with the bias more pronounced at low  $SNR_0$  since the distribution of  $T_1$  is shifted more to the right when the  $M_0/\sigma$  ratio is lower (group B in Fig. 1a). The precision of estimated  $T_1$  from GLLS and NLS is similar since the distributions under the same  $SNR_0$  are similar. Figure 1b shows the bias as a function of  $T_1$  and  $SNR_0$ . For a given  $SNR_0$ , the relative error of  $T_1$  is found to be proportional to the value of  $T_1$ , although the accuracy of  $T_1$  is relatively unaffected by the value of  $T_1$  in a high  $SNR_0$  regime ( $>200$ ). When given the same  $T_1$ , the relative error of  $T_1$  was found to be inversely proportional to  $SNR_0$ . The precision of estimated  $T_1$  from GLLS and NLS is also similar (data not shown).

The results in Fig. 2 show that the WLLS virtually eliminates the  $T_1$  bias for a broad range of  $SNR_0$  and  $T_1$  values. Figure 2a shows that WLLS and NLS produce comparable accuracy of  $T_1$  at all  $SNR_0$ 's tested, while GLLS overestimates  $T_1$  progressively as  $SNR_0$  decreases. Figure 2b shows that the bias of  $T_1$  is corrected in WLLS with the relative error less than 5% regardless of the  $T_1$  value.  $T_1$ 's estimated using WLLS and NLS have similar precision;  $T_1$  estimated using GLLS has slightly lower precision than  $T_1$  estimated using NLS, in agreement with previous reports (10,12) (data not shown).

Figure 3 shows maps of the relative error on the estimated  $T_1$  in the synthetic human brain data using the

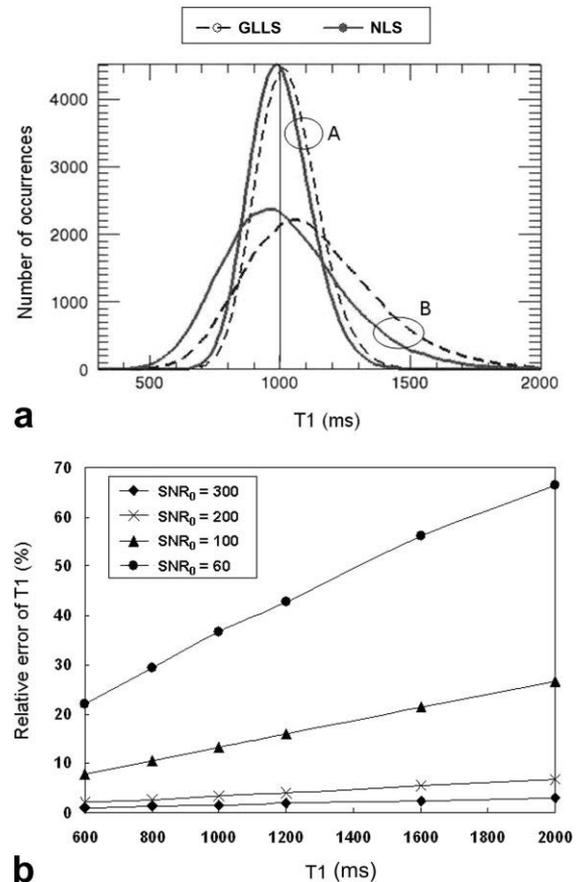


FIG. 1. (a) Distribution of  $T_1$  using six SPGR images (three replicates of two flip angles) with two different noise levels (group A:  $M_0/\sigma = 200$ , group B:  $M_0/\sigma = 100$ ). The true  $T_1$  value is 1000 ms. (b) Relative errors of  $T_1$  using the GLLS approach at different noise levels ( $SNR_0 = 60, 100, 200$ , and  $300$ ) with the true  $T_1$  value set to 600, 800, 1000, 1200, 1600, and 2000 ms. Relative error of  $T_1 = 100 \times (\text{Estimated } T_1 - \text{True } T_1)/\text{True } T_1$ .

GLLS, ILLS, and WLLS methods. The results shown in Fig. 3 were scaled in the range of negative/positive 20% and the gray background corresponds to zero. The relative error of GLLS is consistently higher than that of WLLS in the brain tissue, and is positive, indicating that  $T_1$  is overestimated. The error of ILLS is higher than the error of WLLS in most of the brain tissue. Although the error of ILLS is generally lower than that of GLLS, it shows a strong dependency on  $T_1$ , resulting in an undesirable tissue-dependent pattern. It is also notable that the bias in the cerebral spinal fluid (CSF) regions is higher than the bias in the brain parenchyma for all three methods, with GLLS and WLLS showing positive bias and ILLS negative bias.

## DISCUSSION AND CONCLUSIONS

The linear model for  $T_1$  estimation presented by Gupta (9) and used in many previous works (7–12) is an example of linearization. In general, weighted uncertainties must be used with linearly transformed data because the transformation distorts the experimental errors (16). However, Gupta’s linear regression assumes that the scatter of points

around the line follows a Gaussian distribution and that the SD is the same at every data point. In this work we show that neglecting such adjustments to the uncertainty produces significant errors in the  $T_1$  estimation. The magnitude of  $T_1$  bias can be related to the true  $T_1$  value, and the experiment design affecting SNR, such as the TR, flip angles, image resolution, and receiver coil. For clinical whole-brain SPGR data acquired at 1.5T with a single-channel receiver coil (TR = 8 ms, 1 mm<sup>3</sup> resolution, and flip angles = 2°, 3°, 14°, and 17°), the  $SNR_0$  ranges from 100 to 200 in brain tissue; the  $T_1$  value can be overestimated by 10–20%.

In many applications, achieving an unbiased estimation of the desired parameters from transformed data is achieved by computing weighted uncertainties with error analysis and error propagation techniques. We show such an approach in the Appendix. Moreover, we have derived a new WLLS model for  $T_1$  estimation directly from the nonlinear model without using any transformation or approximation. This is an interesting result because converting a nonlinear model to a linear one using direct analytical derivation without approximations is not always possible.

It is worth emphasizing that the bias of the linear model depends not only on the measurement errors in the abscissa and the ordinate, but also on their covariance since

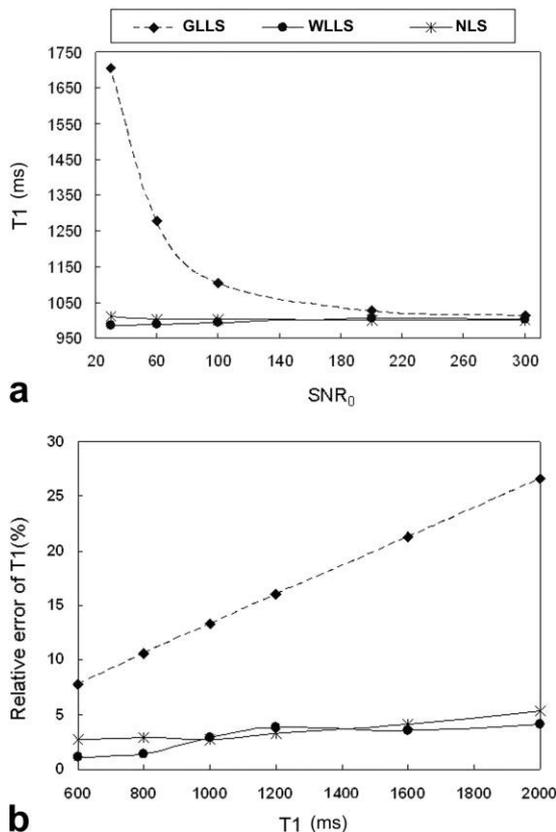


FIG. 2. (a) Estimated  $T_1$  using GLLS, WLLC, and NLS methods assuming a true  $T_1$  value of 1000 ms, and (b) relative error of  $T_1$  using GLLS, WLLS, and NLS methods with  $SNR_0 = 100$ . Six SPGR images, consisting of three replicates of two flip angles without averaging, were used in both a and b.

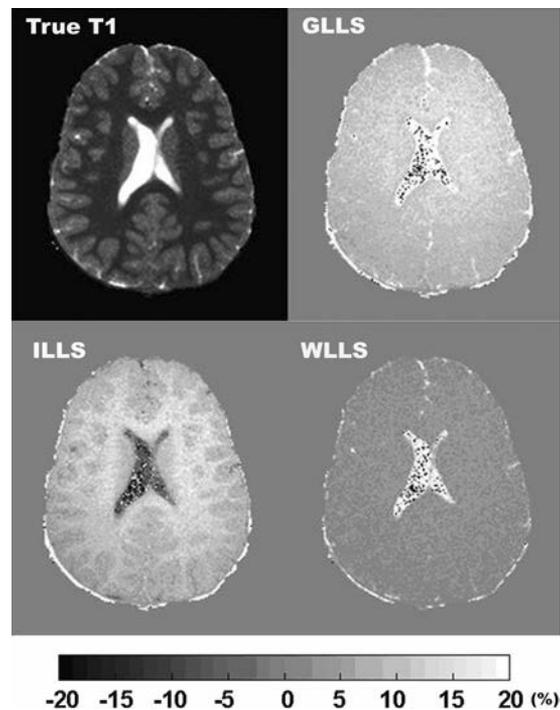


FIG. 3. Relative error of  $T_1$  on a selected slice of synthetic human brain data using the GLLS, WLLS, and ILLS methods in the fitting procedure. The true  $T_1$  map of the same slice is shown in the upper left panel for reference.

the measurements in both axes are no longer independent. Our simulations show that using correct weights on the abscissa and the ordinate in Gupta’s formula will not result in unbiased estimates of  $T_1$ ; a covariance term must be included (see Appendix).

The previously proposed intensity-based weighting approach, ILLS, improves the accuracy of the estimated  $T_1$  in some brain areas. However, with this method the bias shows an undesirable strong dependence on the values of  $T_1$ . In general, all linear methods—GLLS, ILLS, and WLLS—are least accurate in estimating  $T_1$  in regions with high  $T_1$ , such as CSF, but GLLS and WLLS overestimate while ILLS underestimates. This poor performance in CSF has little practical relevance because accuracy in CSF is generally biologically less important than in brain parenchyma. Moreover, the estimation of  $T_1$  from SPGR data in regions of high  $T_1$ , such as CSF, is already problematic given that the SD of  $T_1$  measured from SPGR signals is proportional to the square of  $T_1$  (10). The SPGR simulations we performed in the human brain used experimental parameters aimed at optimizing typical values of  $T_1$  in gray ( $T_1 \sim 950$  ms) and white ( $T_1 \sim 600$  ms) matter, but not in CSF ( $T_1 \sim 4500$  ms), resulting in a magnification of potential problems in regions with high  $T_1$ . Although the results of the WLLS method may be affected by rectified noise, which is not accounted for in all models previously studied, signal correction methods such as those of Koay et al. (15) and Henkelman (24) may be used to resolve this problem. For scanning protocols used in clinical applications, the SNR in brain parenchyma is generally high enough to avoid effects from rectified noise.

$T_1$  bias can also originate from sources that are not addressed here. For example, flip angle variations caused by  $B_1$  inhomogeneity can cause additional errors in  $T_1$  estimation; approaches have been proposed to correct these inaccuracies (12,25). To get a robust estimation of  $T_1$  from clinical SPGR signals, correcting  $B_1$  inhomogeneity or the flip angles should be considered in addition to using the proposed weighted linear method in the fitting.

In terms of computation speed, WLLS is slower than GLLS since WLLS may need several iterations of linear regression while GLLS needs only one; however, WLLS is still computationally faster than NLS. For example, the numerical simulation (Fig. 2) we performed took 11–12 s for GLLS and 59–65 s for WLLS, but 17–29 min for NLS. The difference in time for the same method varied due to different noise levels.

WLLS and NLS are comparable in terms of both precision and accuracy in estimating  $T_1$  and  $M_0$ . WLLS, however, was found to be more stable than NLS at low  $SNR_0$  (i.e., a lower occurrence of  $T_1$  outliers such as negative  $T_1$  values). We suspect that this instability of the NLS approach is due to the known large-residuals problem in nonlinear regression (26). The Newton method for nonlinear fitting is known to be more robust for noisy data than the Levenberg-Marquardt based approaches (18). In future experiments we plan to systematically compare WLLS and NLS using the Newton method in testing the instability in the very low SNR regime. In general, clinical SPGR signals have higher  $SNR_0$  and do not have the problem described above.

In this work we have shown that the relaxation time,  $T_1$ , estimated from the SPGR signals using the widely accepted LLS model is biased. The bias stems from neglecting to adjust uncertainties when transforming a nonlinear model into a linear one. We propose a weighting approach for the linear model that can be derived from the nonlinear model without any approximation. The proposed WLLS method yields estimated  $T_1$  with precision and accuracy comparable to that obtained from nonlinear fitting while reducing the computation time significantly, enabling the generation of robust  $T_1$  maps at the scanner console.

## APPENDIX

Nonlinear regression is generally done without weighting or with constant weighting for all experimental data. For example, the  $\sigma_i$  values in Eq. [2] are the same for all SPGR data points. Giving equal weight to all data points is appropriate when the experimental uncertainty is expected to be the same in all measurements. When transforming a nonlinear function into a linear function, however, we must use weighted (or adjusted) uncertainties  $\sigma_i$  instead of  $\sigma$ , to account for the transformation of the dependent variables (16). In general, if we fit the function  $f(u)$  rather than  $u$ , the uncertainties in the measured quantities must be modified using the following formula (16):

$$\sigma_i = \sigma_{f(u_i)} = \frac{\partial f(u_i)}{\partial u_i} \sigma_i. \quad [A1]$$

Note that the uncertainties should be modified in both abscissa and ordinate in the linear form of LLS (Eq. [3])

since both  $y_i$  and  $x_i$  are now subject to measurement errors on signals,  $s_i$ . Therefore, we have

$$\sigma_{y_i}^2 = \left( \frac{\partial f(s_i)}{\partial s_i} \right)^2 \sigma_i^2 = \left( \frac{1}{\sin(\alpha_i)} \right)^2 \sigma^2 \quad [A2]$$

$$\sigma_{x_i}^2 = \left( \frac{\partial g(s_i)}{\partial s_i} \right)^2 \sigma_i^2 = \left( \frac{1}{\tan(\alpha_i)} \right)^2 \sigma^2. \quad [A3]$$

Also note that the measurement errors due to noise in the abscissa and the ordinate are not independent; therefore, the covariance term between the measurement errors of  $y_i$  and  $x_i$  should also be taken into account:

$$\sigma_{x_i y_i}^2 = \frac{\partial f(s_i)}{\partial s_i} \times \frac{\partial g(s_i)}{\partial s_i} \times \sigma_i^2 = \left( \frac{1}{\sin(\alpha_i)} \right) \left( \frac{1}{\tan(\alpha_i)} \right) \sigma^2. \quad [A4]$$

When applying the correct weighted uncertainties to Gupta's linear model—which we denote as WLLS—using the error propagation equation described in Refs. 16 and 25, we use the new variance that takes the uncertainties in both  $y_i$  and  $x_i$ , and their covariance term into account:

$$\begin{aligned} Var(\sigma_i) &= Var(y_i - bx_i - a) \\ &= \left( \frac{1}{\sin(\alpha_i)} \right)^2 \sigma_i^2 + b^2 \left( \frac{1}{\tan(\alpha_i)} \right)^2 \sigma_i^2 \\ &\quad - 2b \left( \frac{1}{\sin(\alpha_i)} \right) \left( \frac{1}{\tan(\alpha_i)} \right) \sigma_i^2 \\ &= \left( \frac{1 - b \cos(\alpha_i)}{\sin(\alpha_i)} \right)^2 \sigma_i^2 \end{aligned} \quad [A5]$$

The transformed linear equation therefore has the following  $\chi^2$  objective function (16,17,20):

$$\chi_{WLLS}^2(a, b) = \sum_{i=1}^n w(i) x(y_i - a - bx_i)^2, \quad [A6]$$

where  $w_{WLLS}(i) = \frac{1}{Var(\sigma_i)} = \frac{1}{\sigma_i^2} \left( \frac{\sin(\alpha_i)}{1 - b \cos(\alpha_i)} \right)^2$

Note that  $\sigma_i = \sigma$  is a constant and therefore can be factored out.

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## REFERENCES

1. Detre JA, Leigh JS, Williams DS, Koretsky AP. Perfusion imaging. *Magn Reson Med* 1992;23:37–45.
2. Gowland P, Mansfield P, Bullock P, Stehling M, Worthington B, Firth J. Dynamic studies of gadolinium uptake in brain tumors using inversion-recovery echo-planar imaging. *Magn Reson Med* 1992;26:241–258.

3. Vymazal J, Righini A, Brooks RA, Canesi M, Mariani C, Leonardi M, Pezzoli G. T1 and T2 in the brain of healthy subjects, patients with Parkinson disease, and patients with multiple system atrophy: relation to iron content. *Radiology* 1999;211:489–495.
4. Williamson P, Pelz D, Merskey H, Morrison S, Karlik S, Drost D, Carr T, Conlon P. Frontal, temporal, and striatal proton relaxation times in schizophrenic patients and normal comparison subjects. *Am J Psychiatry* 1992;149:549–551.
5. Larsson HB, Frederiksen J, Petersen J, Nordenbo A, Zeeberg I, Henriksen O, Olesen J. Assessment of demyelination, edema, and gliosis by in vivo determination of T1 and T2 in the brain of patients with acute attack of multiple sclerosis. *Magn Reson Med* 1989;11:337–348.
6. Zhang H, Shea SM, Park V, Li D, Woodard PK, Gropler RJ, Zheng J. Accurate myocardial T1 measurements: toward quantification of myocardial blood flow with arterial spin labeling. *Magn Reson Med* 2005;53:1135–1142.
7. Deoni SC, Rutt BK, Peters TM. Rapid combined T1 and T2 mapping using gradient recalled acquisition in the steady state. *Magn Reson Med* 2003;49:515–526.
8. Deoni SC, Peters TM, Rutt BK. High-resolution T1 and T2 mapping of the brain in a clinically acceptable time with DESPOT1 and DESPOT2. *Magn Reson Med* 2005;53:237–241.
9. Gupta R. A new look at the method of variable nutation angle for the measurement of spin-lattice relaxation time using Fourier transform NMR. *J Magn Reson* 1977;25:231–235.
10. Wang HZ, Riederer SJ, Lee JN. Optimizing the precision in T1 relaxation estimation using limited flip angles. *Magn Reson Med* 1987;5:399–416.
11. Fram EK, Herfkens RJ, Johnson GA, Glover GH, Karis JP, Shimakawa A, Perkins TG, Pelc NJ. Rapid calculation of T1 using variable flip angle gradient refocused imaging. *Magn Reson Imaging* 1987;5:201–208.
12. Cheng HL, Wright GA. Rapid high-resolution T1 mapping by variable flip angles: accurate and precise measurements in the presence of radiofrequency field inhomogeneity. *Magn Reson Med* 2006;55:566–574.
13. Chang LC, Koay CG, Deoni S, Pierpaoli C. Comparison of linear and non-linear fitting methods for estimating T1 from SPGR signals. In: *Proceedings of the 15th Annual Meeting of ISMRM, Berlin, Germany, 2007 (Abstract 1775)*.
14. Deoni SC, Peters TM, Rutt BK. Determination of optimal angles for variable nutation proton magnetic spin-lattice, T1, and spin-spin, T2, relaxation times measurement. *Magn Reson Med* 2004;51:194–199.
15. Koay CG, Basser PJ. Analytically exact correction scheme for signal extraction from noisy magnitude MR signals. *J Magn Reson* 2006;179:317–322.
16. Bevington P. *Data reduction and error analysis for the physical sciences*. New York: McGraw-Hill Book Company; 1969.
17. Press WH, Teukolsky SA, Vetterling WT, Flannery BP. *Numerical recipes in C*. Cambridge, UK: Cambridge University Press; 1992. p 666–670.
18. Koay CG, Chang LC, Carew JD, Pierpaoli C, Basser PJ. A unifying theoretical and algorithmic framework for least squares methods of estimation in diffusion tensor imaging. *J Magn Reson* 2006;182:115–125.
19. Chang LC, Jones DK, Pierpaoli C. RESTORE: robust estimation of tensors by outlier rejection. *Magn Reson Med* 2005;53:1088–1095.
20. Barker DR, Diana LM. Simple method for fitting data when both variables have uncertainties. *Am J Phys* 1974;42:224–227.
21. Reed BC. Linear least-squares fits with errors in both coordinates. *Am J Phys* 1989;57:642–646.
22. Brent R. *Algorithms for minimization without derivatives*. Englewood Cliffs, NJ: Prentice-Hall; 1973.
23. Press WH, Teukolsky SA, Vetterling WT, Flannery BP. *Numerical recipes in C*. Cambridge, UK: Cambridge University Press; 1992. p 402–405.
24. Henkelman RM. Measurement of signal intensities in the presence of noise in MR images. *Med Phys* 1985;12:232–233.
25. Deichmann R, Haase A. Quantification of T1 values by Snapshot-FLASH NMR imaging. *J Magn Reson* 1992;96:608–612.
26. Jorge N, Wright SJ. *Numerical optimization*. New York: Springer-Verlag, Inc.; 1999.